



Treatment failure and drug resistance in natural *Leishmania* populations: A multi-disciplinary research consortium

J.C. Dujardin, S. Decuyper, S. De Doncker, M. Boelaert, V. Yardley, S. Croft,
S. Rijal, R. Singh, B. Khanal, J. Arevalo, A. Llanos-Cuentas, H. Bermudez, A. Maurer, F. Chappuis
ITMA, Belgium; LSHTM, UK; BPKIHS, Nepal; IMTA vH, Peru; CUMETROP, Bolivia; HUG, Switzerland

- Control of leishmaniasis is challenged by increasing antimonial (SbV) treatment failure. This can be due to the host, the drug and/or the parasite. Parasite resistance is a well-established fact and several mechanisms were reported in **artificially** induced resistant strains. It is unknown if they are valid in naturally resistant strains.
- We aimed to understand the phenomenon of treatment failure in a global and integrated way and particularly to tackle the genetic bases of parasite resistance in **natural** populations.
- We achieved our objectives through a unique multi-disciplinary study, combining:

Clinical work. 282 Patients with VL (Nepal), 746 with CL (Peru & Bolivia). Treatment failure: 11,3 % in Nepal, 23,9 % in Peru. 562 fully documented strains (110 from treatment failure); identification of clinical risk factors for treatment failure.

Characterisation of isolates.

SbV Susceptibility assays (99 isolates) and species typing

- a) 47 resistant, 52 sensitive
- b) SbV resistance identified in *L. donovani*, *L. braziliensis*, *L. guyanensis*, *L. lainsoni*
- c) No cross-Resistance with miltefosine

Putative SbV resistance markers.

Resistance as a stepwise process (SbV then SbIII)?; upregulation of genes encoding oxidative stress protective proteins; Q-PCR assay for promastigotes, good predictive value for treatment outcome

Immunology.

Cytokine measurement *in situ* (CL), LST

High level of IL-13 is linked to poor response to treatment. LST induration size smaller in treatment failure

Field Work

Parasite Population Genetics.

Effect of transmission (zoo- or anthroponotic) on spreading resistance? Result *L. donovani*: Polyclonal structure of the resistant parasite populations; independent events leading to emergence of drug resistance?

General data processing. Low correlation between treatment outcome and *in vitro* drug susceptibility: need to upgrade *in vitro* assays!!!

Prospects:

- Validation of markers on our collection of strains and clinical samples
- Proteomics and complementation genomics
- Definition of reference strains, dissemination of standardised methods
- Recommendations for further use of antimonials





Bibliography

Clinical work: **1.** Bermúdez H. et al. (2006) Efficacy and safety of a generic sodium stibogluconate for the treatment of tegumentary leishmaniasis in Isiboro Secure Park, Bolivia. *Ann.Trop.Med.Hyg.*, 100: 591-600. **2.** Llanos-Cuentas, A. et al. (2007) Clinical and parasite species risk factors for pentavalent antimonial treatment failure in cutaneous leishmaniasis in Peru. *Clinical and Infectious Diseases*, in press. **3.** Rijal, S. et al. (2007) Clinical risk factors for pentavalent antimonial treatment failure in Kala-Azar in Nepal. Final stage.

Immunology: **4.** Maurer-Cecchini, A. et al. (2007) Immunological determinants of clinical outcome in Peruvian patients with tegumentary leishmaniasis treated by pentavalent antimonials. Submitted

Species and treatment outcome: **5.** Arevalo, J. et al. (2007). The influence of *Leishmania (Viannia)* species on the response to antimonial treatment of patients with American Tegumentary Leishmaniasis. *JID*, 195: 1846-51. **6.** Garcia, A.L. et al. (2004). Culture-independent species typing of Neotropical *Leishmania*: clinical validation of a PCR-based assay targeting heat-shock protein 70 genes. *J.Clin.Microb.*, 42: 2294-2297. **7.** Garcia, A.L. et al. (2005) American tegumentary leishmaniasis: antigen-gene polymorphism, taxonomy and clinical pleomorphism. *Infection, Genetics and Evolution*, 5: 109-116. **8.** Garcia, A.L. et al. (2006) American tegumentary leishmaniasis: direct species identification of *Leishmania* in non-invasive clinical samples. *Trans.Roy.Soc.Trop.Med.Hyg.*, 101: 368-71

Drug susceptibility of parasites: **9.** Yardley, V. et al. (2006) American tegumentary leishmaniasis: is antimonial treatment outcome related with parasite drug susceptibility? *JID*, 194(8):1168-75. **10.** Rijal, S. et al. (2007) Antimonial treatment of visceral leishmaniasis: are current in vitro susceptibility assays adequate for prognosis of in vivo therapy outcome? *Microbes and Infection*, 9: 528-35 **11.** Yardley, V. et al. (2005). The sensitivity of clinical isolates of *Leishmania* from Peru and Nepal to miltefosine. *Am.J.Trop.Med.Hyg.*, 73: 272-275.

Mechanisms of drug resistance: **12.** Decuyper, S. et al. (2005) Differential polyadenylation of ribosomal RNA during post-transcriptional processing in *Leishmania*. *Parasitology*, 131: 321-29. **13.** Decuyper, S. et al. (2005). A study of the mechanism of natural Sb(V) resistance in Nepalese *Leishmania donovani* isolates based on gene expression analysis. *AAC*, 49: 4616-21 **14.** Decuyper, S. et al. (2007) Gene expression profiling in *Leishmania*: overcoming technical variation and exploiting biological variation. *Parasitology*, Oct 12 Epub. **15.** Decuyper, S. et al. (2007) Gene expression profiling of clinical *Leishmania donovani* isolates: molecular characterisation yielding biological and epidemiological insights into natural drug resistance and treatment failure. Final stage. **16.** Adai, V. et al. (2007) Gene expression profiling in clinical isolates of *Leishmania braziliensis* differing in antimonial susceptibility. Final stage.

Parasite population genetics: **17.** Laurent, T. et al. (2007) Epidemiological dynamics of antimonial resistance in *Leishmania donovani*: genotyping reveals a polyclonal population structure among naturally-resistant clinical isolates from Nepal. *Infection, Genetics and Evolution*, 7: 206-212. **18.** Adai, V. et al. (2007) Epidemiological dynamics of antimonial resistance in *Leishmania braziliensis* clinical isolates from Peru. Final stage.

Reviews: **19.** Decuyper, S. (2007) Antimonial treatment failure in anthroponotic visceral leishmaniasis: towards improved tools and strategies for epidemiological surveillance and disease control. PhD thesis http://www.itg.be/itg/PhD_Decuyper_E_version.zip **20.** Reithinger, R. and Dujardin, J.C. (2007) Molecular Diagnosis of Leishmaniasis: Current Status and Future Applications. *Journal of Clinical Microbiology*, 45(1):21-5 **21.** Reithinger, R., Dujardin, J.C. et al. (2007) Cutaneous Leishmaniasis: Burden of Disease, Clinical Pathology, Epidemiology, Prevention and Control. *Lancet-Infectious Diseases*, 7: 581-596



Impact

Recommendations (written in february 2006) should be adapted with care, by endemic area.

In Nepal, it is crucial that suspected VL patients are referred early to the district hospitals and centers for confirmation of the diagnosis and early initiation of therapy. It is also important to ensure that the patients are monitored closely so that they complete the therapy. A mechanism to follow the patient in the nearest health facility at the end of therapy and at 6 months follow up should be made to record the clinical outcome of therapy. All kala-azar cases treated at non government health facilities including private facilities should be reported to the district public health office. In view of the increasing failure rates of SSG in kala-azar patients from the districts bordering the Bihar high resistance it would be preferable to treat patients with advanced disease from these areas with Amphotericin B in order to decrease fatalities. We recommend a further assessment of the efficacy of antimonials in districts bordering the focus of high antimonial resistance in Bihar. If a low efficacy is confirmed in these districts, a change of first line therapy should be considered (**Note: since the writing of these recommendations, Miltefosine has been recommended to be the 1st line in the elimination campaign. However SSG continues to be used in Nepal and supplied by the MOH as they plan to phase it out gradually**). Based on our finding that drug resistance is likely to emerge several times through independent events, we also recommend the implementation of regional networks for monitoring of drug efficacy and emergence of drug resistance. These could use the markers and technologies we have developed after their further validation.

In Peru, considering the species-specific differences in treatment outcome, organisations involved in leishmaniasis control should consider treatment strategies adapted to the parasites circulating in the region. This might have a direct economic implication. New therapeutic schemes are necessary and particularly adapted to children; combination of treatment should be considered, involving immuno-modulatory drugs. Diagnostic procedures should be improved and standardised, whilst follow-up strategies should be implemented for the early detection of treatment failure. Several clinical features have been identified as significant predictors of treatment failure in CL patients. Further work to develop clinical scores predicting treatment failures should be conducted.